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(54) Title: SACCHARIDE COMPOSITIONS, METHODS AND APPARATUS FOR THEIR SYNTHESIS (57) Abstract A method for preparing saccharide compositions is disclosed. The method is reiterative and comprises the following three steps: (i) a glycosyltransferase capable of transferring a preselected saccharide unit to an acceptor moiety is isolated by contacting the acceptor moiety with a mixture suspected of containing the glycosyltransferase under conditions effective to bind the acceptor moiety and the glycosyltransferase and thereby isolate the glycosyltransferase. The acceptor moiety is a protein, a glycoprotein, a lipid, a glycolipid, or a carbohydrate. (ii) The isolated glycosyltransferase is then used to catalyze the bond between the acceptor moiety and the preselected saccharide unit. (iii) Steps (i) and (ii) are repeated a plurality of times with the intermediate product obtained in the first iteration of the method being used as the acceptor moiety of the second iteration.		

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DescriptionSaccharide Compositions, Methods and
Apparatus for their SynthesisTechnical Field

5 This invention relates to saccharide compositions such as, for example, oligosaccharides, polysaccharides, glycolipids, and glycoproteins. More specifically, this invention relates to processes for preparing these and other saccharide compositions by enzymatic techniques.

Background Art

10 The term "carbohydrate" embraces a wide variety of chemical compounds having the general formula $(CH_2O)_n$, such as monosaccharides, disaccharides, oligosaccharides and polysaccharides. Oligosaccharides are chains
15 composed of saccharide units, which are alternatively known as sugars. These saccharide units can be arranged in any order and the linkage between two saccharide units can occur in any of approximately ten different ways. As a result, the number of different possible stereoisomeric
20 oligosaccharide chains is enormous.

 Of all the biological polymer families, oligosaccharides and polysaccharides have been the least well studied, due in considerable part to the difficulty of sequencing and synthesizing their often complex sugar
25 chains. Although the syntheses of oligonucleotides and polypeptides are well developed, there is currently no generally applicable synthetic technique for synthesizing oligosaccharides.

 Numerous classical techniques for the synthesis of
30 carbohydrates have been developed, but these techniques suffer the difficulty of requiring selective protection and deprotection. Organic synthesis of oligosaccharides is further hampered by the lability of many glycosidic

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bonds, difficulties in achieving regioselective sugar coupling, and generally low synthetic yields. These difficulties, together with the difficulties of isolating and purifying carbohydrates and of analyzing their structures, has made this area of chemistry a most demanding one.

Much research effort has been devoted to carbohydrates and molecules comprising carbohydrate fragments, such as glycolipids and glycoproteins. Research interest in such moieties has been largely due to the recognition that interactions between proteins and carbohydrates are involved in a wide array of biological recognition events, including fertilization, molecular targeting, intercellular recognition, and viral, bacterial, and fungal pathogenesis. It is now widely appreciated that the oligosaccharide portions of glycoproteins and glycolipids mediate recognition between cells and cells, between cells and ligands, between cells and the extracellular matrix, and between cells and pathogens.

These recognition phenomena can likely be inhibited by oligosaccharides having the same sugar sequence and stereochemistry found on the active portion of a glycoprotein or glycolipid involved in cell recognition. The oligosaccharides are believed to compete with the glycoproteins and glycolipids for binding sites on receptor proteins. For example, the disaccharide galactosyl B 1-4 N-acetylglucosamine is believed to be one component of the glycoproteins which interact with receptors in the plasma membrane of liver cell. Thus, to the extent that they compete with potentially harmful moieties for cellular binding sites, oligosaccharides and other saccharide compositions have the potential to open

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new horizons in pharmacology, diagnosis, and therapeutics.

There has been relatively little effort to test oligosaccharides as therapeutic agents for human or animal diseases, however, as methods for the synthesis of oligosaccharides have been unavailable as noted above. Limited types of small oligosaccharides can be custom-synthesized by organic chemical methods, but the cost for such compounds is typically very high. In addition, it is very difficult to synthesize oligosaccharides stereospecifically and the addition of some sugars, such as sialic acid and fucose, has not been effectively accomplished because of the extreme lability of their bonds. Improved, generally applicable methods for oligosaccharide synthesis are desired for the production of large amounts of widely varying oligosaccharides for pharmacology and therapeutics.

For certain applications, enzymes have been targeted for use in organic synthesis as one alternative to more traditional techniques. For example, enzymes have been used as catalysts in organic synthesis; the value of synthetic enzymatic reactions in such areas as rate acceleration and stereoselectivity has been demonstrated. Additionally, techniques are now available for low cost production of some enzymes and for alteration of their properties.

The use of enzymes as catalysts for the synthesis of carbohydrates has been proposed, but to date enzyme-based techniques have not been found which are useful for the general synthesis of oligosaccharides and other complex carbohydrates in significant amounts. It has been

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recognized that a major limiting factor to the use of enzymes as catalysts in carbohydrate synthesis is the very limited current availability of the broad range of enzymes required to accomplish carbohydrate synthesis.

- 5 See Toone et al, Tetrahedron Reports (1990) (45)17:5365-5422.

In mammalian systems, eight monosaccharides activated in the form of nucleoside mono- and diphosphate sugars provide the building blocks for most
10 oligosaccharides: UDP-Glc, UDP-GlcUA, UDP-GlcNAc, UDP-Gal, UDP-GalNAc, GGP-Man, GDP-Fuc and CMP-NeuAc. These are the intermediates of the Leloir pathway. A much larger number of sugars (e.g., xylose, arabinose) and oligosaccharides are present in microorganisms and
15 plants.

Two groups of enzymes are associated with the in vivo synthesis of oligosaccharides. The enzymes of the Leloir pathway is the largest group. These enzymes transfer sugars activated as sugar nucleoside phosphates
20 to a growing oligosaccharide chain. Non-Leloir pathway enzymes transfer carbohydrate units activated as sugar phosphates, but not as sugar nucleoside phosphates.

Two strategies have been proposed for the enzyme-catalyzed in vitro synthesis of oligosaccharides. See
25 Toone et al, supra. The first strategy uses glycosyltransferases. The second uses glycosidases or glycosyl hydrolases.

Glycosyltransferases catalyze the addition of activated sugars, in a stepwise fashion, to a protein or
30 lipid or to the non-reducing end of a growing oligosaccharide. A very large number of

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glycosyltransferases appear to be necessary to synthesize carbohydrates. Each NDP-sugar residue requires a distinct class of glycosyltransferases and each of the more than one hundred glycosyltransferases identified to date appears to catalyze the formation of a unique glycidic linkage. To date, the exact details of the specificity of the glycosyltransferases are not known. It is not clear, for example, what sequence of carbohydrates is recognized by most of these enzymes.

10 Enzymes of the Leloir pathway have begun to find application to the synthesis of oligosaccharides. Two elements are required for the success of such an approach. The sugar nucleoside phosphate must be available at practical cost and the glycosyltransferase must be available. The first issue is resolved for most common NDP-sugars, including those important in mammalian biosynthesis. The problem in this technology however resides with the second issue. To date, only a very small number of glycosyltransferases are available.

20 Access to these types of enzymes has been the single limiting factor to this type of carbohydrate synthesis.

It has been reported that most glycosyltransferases are difficult to isolate, particularly from mammalian source. This is because these proteins are present in low concentrations and are membrane-bound. Further, although a few glycosyltransferases have been immobilized, these enzymes have been reported to be unstable. To date only a very small number of glycosyltransferases are available from commercial sources, and these materials are expensive.

Much hope has therefore been put on future developments in genetic engineering (i.e., cloning) of

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enzymes, particularly since several glycosyltransferases have already been cloned, including galacto-, fucosyl-, and sialyltransferases. It is hoped that future advances in cloning techniques will speed the cloning of other glycosyltransferases and enhance their stability.

Accordingly, in light of their potential uses and the difficulty or impossibility to obtain them in sufficient quantities, there exists a long-felt need for general synthetic methods for the production of oligosaccharides, polysaccharides, glycoproteins, glycolipids, and similar species in an efficient, cost effective, stereospecific, and generally applicable manner.

Disclosure of the Invention

It is an object of the present invention to provide saccharide compositions, particularly oligosaccharides, polysaccharides and chemical moieties which comprise oligosaccharide units.

It is another object of this invention to provide a wide variety of saccharide compositions, including those not found in nature.

It is a further object of this invention to provide saccharide compositions useful in mitigating the effects of human or animal diseases.

It is yet another object of this invention to provide improved processes for preparing saccharide compositions.

It is a further object of this invention to provide enzymatic processes for preparing saccharide

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compositions.

It is still another object of this invention to provide processes for obtaining enzymes useful in synthesizing saccharide compositions.

5 It is still another object of this invention to provide an apparatus useful for the synthesis of saccharide compositions in accordance with the present invention.

10 These and other objects are achieved by the present invention, which provides enzymatic processes for preparing oligosaccharides, polysaccharides, glycolipids, glycoproteins, and other saccharide compositions. These processes involve the enzyme-facilitated transfer of a
15 preselected saccharide unit from a donor moiety to an acceptor moiety. Saccharide compositions having a plurality of saccharide units are preferably prepared by appending the saccharide units in stepwise fashion to acceptor moieties which are themselves saccharide compositions prepared in accordance with this invention.

20 Accordingly, methods for preparing saccharide compositions are provided comprising the steps of providing an acceptor moiety and contacting the acceptor moiety with a glycosyltransferase. The glycosyltransferase is prepared so as to be specific for
25 the acceptor moiety and capable of transferring a saccharide unit to the acceptor moiety. This method of the present invention is performed a plurality of times such that the product of the first iteration becomes the acceptor moiety for a second iteration, and so forth.

30 Brief Description of the Drawings

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Figures 1, 2 and 3 illustrate apparatuses suitable for use in the glycosyltransferase catalyzed synthesis of saccharide composition in accordance with the invention.

Best Mode for Carrying Out the Invention

5 As employed herein, the term "saccharide composition" is intended to include any chemical moiety having a saccharide unit within its structure. Sugars, carbohydrates, saccharides, monosaccharides, oligosaccharides, polysaccharides, glycoproteins, and
10 glycolipids provide examples of saccharide compositions. Mixtures and solutions comprising such moieties are also saccharide compositions.

 Saccharide compositions are prepared according to this invention by the enzyme facilitated transfer of
15 saccharide units from donor moieties to acceptor moieties. It will be appreciated that such transfer occurs upon contacting the acceptor and donor moieties with a glycosyltransferase, and typically results in covalently bonding of the acceptor moiety and the
20 saccharide unit stereoselectively, that is, in but one stereoisomeric form.

 The saccharide compositions prepared in accordance with this invention are believed to find wide utility in diagnostics, therapeutics, and pharmacological
25 applications. Once the sugar sequence of a desired target saccharide composition has been determined by conventional methods, a retrosynthetic analysis is generally performed to determine an appropriate synthetic scheme for the saccharide composition. Such a synthetic
30 scheme preferably identifies the particular donor moieties, acceptor moieties, and glycosyltransferases necessary to yield the desired saccharide composition.

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Instead of relying on future developments in genetic engineering to provide the numerous glycosyltransferases required for carbohydrate synthesis, the present invention relies on the very different approach as follows. In the synthesis of a saccharide composition in accordance with the invention, a preselected saccharide unit is first enzymatically attached to an initial acceptor moiety, i.e., a protein, a glycoprotein, a lipid, a glycolipid, or a carbohydrate starting material. This is followed by enzymatically attaching preselected saccharide units to the product obtained in a stepwise fashion thereby forming the saccharide composition.

With the attachment of each preselected saccharide unit one obtains an intermediate product. The present invention is based on the inventor's discovery that the starting material of the synthesis (i.e., the protein, glycoprotein, lipid, glycolipid or carbohydrate) and each intermediate product formed in the synthesis can be advantageously used to obtain, for each corresponding step of the synthesis, a glycosyltransferase specific to catalyze the attachment of the next intermediate product in the synthesis of the target saccharide composition.

Thus, in accordance with the invention, the glycosyltransferase needed for any given step is isolated with the intermediate product (the acceptor moiety) and used to attach to the acceptor moiety the next saccharide unit necessary for construction of the target carbohydrate molecule. In accordance with the present invention, this process is repeated, with each iteration (time) yielding the particular glycosyltransferase required to attach the next saccharide unit onto the growing molecule being isolated, until the target

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carbohydrate molecule is obtained.

Also provided by the invention are reaction conditions and co-reagents as may be necessary and sufficient to effect the covalent bonding of the saccharide unit to the acceptor moiety.

In accordance with preferred embodiments, the acceptor moiety may be a protein, glycoprotein, lipid, glycolipid, or carbohydrate, such as a monosaccharide, disaccharide, oligosaccharide, or polysaccharide. In accordance with other preferred embodiments, the glycosyltransferase is attached to a solid support.

The present methods are capable of stereospecific attachment of the saccharide unit to the acceptor moiety. In general, it is preferred to employ saccharide nucleotides as donor moieties. Uridine, guanosine, and cytidine phosphate materials terminated by the saccharide units to be donated preferably comprise the donor moieties.

The present invention thus also provides means for preparing a glycosyltransferase specific for a particular acceptor moiety and capable of transferring a preselected saccharide unit to the acceptor moiety. Such methods comprise contacting the acceptor moiety with a mixture suspected to contain a plurality of glycosyltransferases under conditions effective to bind the acceptor moiety and the glycosyltransferase specific for the acceptor moiety. The resulting, bound glycosyltransferase is subsequently isolated. It is preferred that the glycosyltransferase be sequenced and that the glycosyltransferase be produced in enhanced quantities by genetic engineering techniques.

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The mixture suspected to contain a glycosyltransferase of interest may be identified as follows. For the most common glycosidic linkages, the glycosyltransferase activities have been described in publications. This is largely true for compounds like milk oligosaccharides, or the carbohydrate moieties of typical (i.e., prevalent) glycoproteins and glycolipids. For less well described linkages, one may first look to the tissue, organ, foodstuff organism, in which the linkage is found. Generally, if the linkage is found in a particular source, the enzyme that made the linkage is also present in the source.

If one is presented only with a saccharide structure, and not a source, one can then test examples of organisms that are likely to contain such a saccharide structure using the most sensitive screening assay available. For example, if the compound contained iduronic acid, N-acetylgalactosamine and N-acetylglucosamine, one would test vertebrate connective tissue. If the target compound contain abequose, one would test bacteria and plants for the presence of the appropriate glycosyltransferase.

Various assays for detecting glycosyltransferases which can be used in accordance with the invention have been published. The following are illustrative. Furukawa et al, Biochem. J., (1985) 227:573-582 describe a borate-impregnated paper electrophoresis assay and a fluorescence assay (figure 6) developed by the inventor. Roth et al, Exp'l Cell Research (1983) 143:217-225 describe application of the borate assay to glucuronyl transferases, previously assayed colorimetrically. Benau et al, J. Histochem. Cytochem. (1990) 38(1):23-30 describe a histochemical assay based on the reduction, by

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NADH, of diazonium salts.

Once a source for the glycosyltransferase of interest has been found, the source is homogenized. The enzyme is purified from homogenate by affinity chromatography using the acceptor moiety as the affinity ligand. That is, the homogenate is passed over a solid matrix having immobilized thereon the acceptor moiety under conditions which cause the glycosyltransferase to bind to the acceptor moiety. The solid support matrix having the glycosyltransferase bound thereto is then washed. This is followed by an elution step in which the glycosyltransferase is desorbed from the solid support matrix and collected. As known, the absorbed glycosyltransferase may be eluted, for example, by passing an aqueous salt (e.g. NaCl) solution over the solid support matrix.

In actual practice of the invention, the "enzyme" purified from the homogenate by affinity chromatography and which is used to attack a preselected saccharide unit onto the acceptor moiety comprises a mixture of various glycosyltransferases which have been purified from other extraneous biological material present in the homogenate which includes enzymes which can interfere with the desired activity of the purified glycosyltransferases. Thus, the glycosyltransferases used in accordance with the present invention is frequently a mixture of various "glycosyltransferase". If desired, this material may be further purified with a single purified glycosyltransferase being isolated and used in the process of the present invention, but such further purification is generally not necessary.

In accordance with the present invention, an

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acceptor moiety is provided which is capable of being covalently bound to a preselected saccharide unit. Representative acceptor moieties include proteins, glycoproteins, lipids, glycolipids and carbohydrates. It will be appreciated that acceptor moieties are preferred to the extent that they are present as a structural component of a saccharide composition of interest. For example, in preparing a saccharide composition such as N-acetylneuraminy 2-3 galactosyl 8 1-4 N-acetylglucosamine, preferred acceptor moieties would be N-acetylglucosamine and galactosyl 8 1-4 N-acetylglucosamine. It will likewise be appreciated that where an acceptor moiety is terminated by a saccharide unit, subsequent saccharide units will typically be covalently bound to the nonreducing terminus of the terminal saccharide.

The saccharide unit to be transferred to an acceptor moiety is provided by a donor moiety for the saccharide unit. A donor moiety according to this invention includes the saccharide unit to be transferred and is capable of providing that saccharide unit to the acceptor moiety when contacted by the acceptor moiety and the appropriate glycosyltransferase. Preferred donor moieties are saccharide nucleotides, such as saccharide-terminated uridine phosphates, saccharide-terminated guanosine phosphates, and saccharide-terminated cytidine phosphates.

It will be appreciated that donor moieties are preferred to be capable of readily providing their component saccharide unit to an acceptor moiety when placed in contact therewith and with a glycosyltransferase. For example, uridine diphosphate galactose is preferred for transferring galactose to

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N-acetylglucosamine, while cytidine monophosphate N-acetylneuraminic acid is preferred for transferring N-acetylneuraminic acid, a sialic acid, to galactosyl & 1-4 N-acetylglucosamine.

5 Upon identification of acceptor moieties and donor moieties necessary for the preparation of a saccharide composition, a glycosyltransferase for each acceptor/donor pair should be prepared. Those skilled in the art will appreciate that a glycosyltransferase may be
10 broadly defined as an enzyme which facilitates the transfer of a saccharide unit from one chemical moiety (here defined as a donor) to another (here defined as an acceptor) and which is named phenomenologically according to the saccharide unit it transfers. Thus,
15 galactosyltransferase transfers galactose, while fucosyltransferase transfers fucose.

 Glycosyltransferases according to this invention are those able to effect the transfer of a predetermined saccharide unit to an acceptor moiety.
20 Glycosyltransferases are preferably specific for an acceptor moiety or at least some significant, active, or exposed portion thereof. Specificity is manifested for a glycosyltransferase by its tendency to bind with a particularly sequenced portion of an acceptor moiety when
25 placed in contact or close proximity therewith and to effect the transfer of a particular saccharide unit to that acceptor moiety.

 Currently, glycosyltransferases are available only from natural sources and, as a result, are somewhat
30 limited in number. It will be appreciated that known glycosyltransferases are only capable of effecting
 saccharide unit transfers which are highly specific, both

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in terms of the chemical identity of the saccharide unit transferred and the stereochemistry of its subsequent attachment to the acceptor moiety. For example, it is known that one N-acetylneuraminyltransferase can effect the transfer of N-acetylneuraminic acid to an acceptor moiety bearing only a galactose unit to produce a saccharide composition having an α 2-3 linkage between the N-acetylneuraminic acid unit and the galactose unit.

Thus, the invention permits construction of sugar linkages found in nature. For example, the linkage of galactose α 1-2 to N-acetylneuraminic acid, which has not been found in nature, cannot presently be effected. The methods disclosed herein are, however, applicable to any type of glycosyltransferase which may become available.

While the behavior of a number of glycosyltransferases is known, most glycosyltransferases are currently not fully characterized. The present invention, however, provides methods by which all glycosyltransferases amenable to its practice may be identified and prepared. It has now been found that an acceptor moiety can be used as an affinity chromatographic tool to isolate enzymes that can be used to transfer particular saccharide units and, thus, synthesize other glycosides.

In a preferred embodiment, an acceptor moiety is immobilized as, for example, on a solid support. It will be appreciated that the term "solid support" includes semi-solid supports as well. Once immobilized, the acceptor moiety is contacted with a mixture suspected to contain glycosyltransferases, such as one comprising naturally-occurring cell homogenate. Since an immobilized acceptor moiety will bind an enzyme specific

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for it, this system is then monitored for acceptor-bound enzyme.

Monitoring for acceptor-bound enzyme may be carried out as follows. The cell homogenate is passed over the immobilized acceptor moiety. This may be achieved, for example, by passing the cell homogenate over a column charged with immobilized acceptor moiety. The column is then washed and the amount of protein which passes through the column charged with immobilized acceptor moiety is monitored. When no more protein is detected, an aqueous salt solution eluant is passed through the column to elude the enzyme. The eluant obtained is then assayed for the presence of glycosyltransferase(s). The assays which can be used are noted above, i.e., the methods described by Furukawa et al, Roth et al and Benau et al.

If no binding of the enzyme to the acceptor moiety occurs (i.e., the assay of the eluate fails to reveal the presence of glycosyltransferase(s) therein), then it can be concluded that the mixture did not contain an enzyme specific for the particular acceptor. Other mixtures of, for example, animal and/or plant cell homogenates are then contacted with the acceptor moiety until enzyme binding is observed.

When the acceptor moiety is bound by an enzyme, the species are separated and further studied. In a preferred embodiment, the acceptor and the candidate enzyme are again contacted, this time in the presence of a donor moiety which comprises the saccharide unit desired to be transferred to the exceptor moiety. If such contacting results in the transfer of the saccharide unit to the acceptor, the enzyme is a glycosyltransferase

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useful in the practice of this invention.

It will be appreciated that once the glycosyltransferase is identified, it can be sequenced and/or replicated by techniques well-known to those skilled in the art. For example, replication might be accomplished by recombinant techniques involving the isolation of genetic material coding for the glycosyltransferase and the preparation of an immortal cell line capable of producing the glycosyltransferase. Replication will likely prove desirable for commercial scale production of saccharide compositions in accordance with this invention.

After the glycosyltransferase is identified, it is contacted with the acceptor moiety and donor moiety under conditions sufficient to effect transfer and covalently bonding of the saccharide unit to the acceptor moiety. It will be appreciated that the conditions of, for example, time, temperature, and pH appropriate and optimal for a particular saccharide unit transfer can be determined by one of skill in the art through routine experimentation. Certain co-reagents may also prove useful in effecting such transfer. For example, it is preferred that the acceptor and donor moieties be contacted with the glycosyltransferase in the presence of divalent cations, especially manganese cations such as may be provided by $MnCl_2$.

In a preferred embodiment, the glycosyltransferase is immobilized by attachment to a solid support and the acceptor and donor moieties to be contacted therewith are added thereto. As discussed above, the glycosyltransferase used in accordance with the present invention is frequently a mixture of glycosyltransferases

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containing at least one glycosyltransferase possessing the desired activity, but purified single glycosyltransferases may also be used in accordance with the present invention. In this preferred embodiment, either the mixture of glycosyltransferases or the purified single glycosyltransferase may be immobilized. Alternatively, the glycosyltransferase, donor and acceptor are each provided in solution and contacted as solutes.

A preferred procedure for immobilization of glycosyltransferases -- and of acceptor moieties, where necessary -- is based on the copolymerization in a neutral buffer of a water soluble prepolymer such as poly(acrylamide-co-N-acryloxysuccinimide (PAN), a cross-linking diamine such as triethylenetetramine, and the glycosyltransferase, as disclosed by Pollack *et al.*, J. Am. Chem. Soc. (1980) 102:6324-36. The immobilization of the enzymes on PAN is useful because small amounts of enzyme can be used, high yields of enzyme activity are obtained, and the bond between enzyme and polymer is stable.

More preferred methods of immobilization include immobilization of the glycosyltransferase amino groups onto solid support oxirane groups (see, e.g., Chan *et al.*, Enzyme Eng. (1980) 5:457-460) or onto cyanogen bromide activated "SEPHADEX" or "SEPHAROSE" (Axen *et al.*, Nature (1967) 214:1302-1304).

In a preferred embodiment, the glycosyltransferase is immobilized from a moderately purified composition containing the glycosyltransferase. Extremely pure enzyme preparations (ie, with specific activities in the range of 1 μ mole transferred per μ g protein per minute of

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incubation) are less efficiently immobilized covalently to solid supports, in that the percent derivatization is lower, compared to 10 or 100 times less pure preparations.

5 It will be appreciated that impairment of the active sites of the glycosyltransferase due to immobilization should be avoided. The inventor observed that contaminating enzyme activities tend to disappear during the immobilization process when compared to the
10 glycosyltransferase of interest when the glycosyltransferase is specifically protected during the immobilization process. During the immobilization process the glycosyltransferase may be protected by the cation required by the enzyme, the nucleotide recognized
15 by the enzyme, and the acceptor recognized by the enzyme. For example, a galactosyl transferase may be protected with Mn^{2+} , N-acetylglucosamine and UDP during the immobilization, regardless of which mobilization method is used. In this way, contaminating proteases are not
20 protected in any way during the immobilization process.

Because only the desired glycosyltransferase is protected during the immobilization process, enzymes which interfere with the synthesis of the target saccharide composition, the activity of enzymes that
25 interfere with the synthesis of the target saccharide composition tends to be lost. Examples of interfering enzymes are proteases, which would otherwise attack the desired glycosyltransferase, and glycosidases, which would otherwise attack the product saccharide.

30 As noted above, in accordance with the invention, a saccharide composition prepared by contacting an acceptor moiety with a donor moiety and a glycosyltransferase can,

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in turn, serve as an acceptor moiety for isolating further enzymes and as an acceptor moiety to which subsequent saccharide units may be transferred. The addition of saccharide units to saccharide compositions prepared by such contact is preferred for the synthesis of carbohydrates and saccharide chains having greater than about three saccharide units.

For example, in preparing the trisaccharide N-acetylneuraminy1 α 2-3-galactosyl β 1-4 N-acetylglucosamine, the disaccharide galactosyl β 1-4 N-acetylglucosamine is prepared according to this invention and then employed as an acceptor moiety to which a subsequent unit is added. Those skilled in the art will appreciate that the saccharide units attached to the saccharide compositions of this invention can be the same or different.

The saccharide compositions of this invention find use in an exceedingly wide variety of applications and may be used in the same manner as saccharide compositions available from known sources. It is preferred that the saccharide compositions be employed in therapeutic and preventative treatments for mammals, such as disclosed in U.S. serial Number 07/241,012.

The saccharide compositions of this invention are expected to find use as blocking agents for cell surface receptors in the treatment of numerous diseases of viral, bacterial, or fungal origins, such as pneumonia, candidiasis, urinary tract infections, periodontal disease, and diarrhea. For example, oligosaccharides prepared according to this invention may inhibit the attachment of pathogens such as pneumonia-causing bacteria to mammalian membrane molecules. Such pathogens

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might be incubated with cellular glycoproteins and glycolipids that have been separated by chromatography or electrophoresis. After detecting specific adherence patterns, the target compound could be analyzed and
 5 inhibitory saccharide composition prepared. If either of the complimentary molecules functions through its saccharide component, then specific saccharide compositions should inhibit attachment.

10 The saccharide compositions which can be prepared in accordance with the invention can be used in the following applications:

1. Nutritional supplements:

- infant formulas

(e.g., $\text{fuc} \xrightarrow{\alpha 1-2} \text{gal} \xrightarrow{\beta 1-4} \text{glcNAc} \xrightarrow{\beta 1-3} \text{gal} \xrightarrow{\beta 1-4} \text{glu}$)

- 15
- geriatric formulas
 - special care formulas

2. Antibacterials:

- pneumonia (e.g., $\text{galNAc} \xrightarrow{\beta 1-3/4} \text{gal} \xrightarrow{\beta 1-4} \text{glu}$)

- urinary tract infection

20 (e.g., $\text{fuc} \xrightarrow{\alpha 1-2} \text{gal} \xrightarrow{\beta 1-4} \text{glcNAc} \xrightarrow{\beta 1-3} \text{gal} \xrightarrow{\beta 1-4} \text{glu}$)

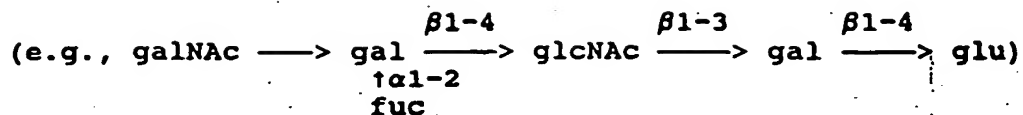
(e.g., dental $\text{gal} \xrightarrow{\alpha 1-4} \text{glu} \xrightarrow{\alpha 1-4} \text{glu}$)₄₋₂₅

- periodontal disease

(e.g., $\text{NAN} \xrightarrow{\alpha 2-3} \text{gal} \xrightarrow{\beta 1-3} \text{galNAc}$)

- 25
- diarrhea

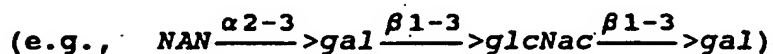
-22-



- 5 - surgical (nosocomial) infections
 - catheter-associated infections

3. Antitumor:

- solid tumor metastases



10

$$\uparrow \alpha 1-4 \\ \text{fuc}$$

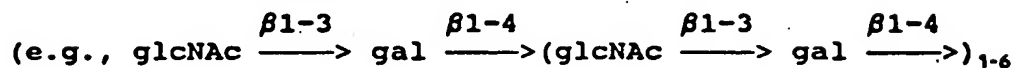
4. Anti-inflammatory:

- neutrophil-platelet interactions
 - WBC-endothelium interactions

15 5. Naval drag-reduction:

- ship hulls

6. Contraceptives



20

- foam and jelly components

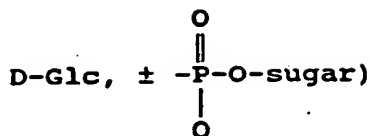
7. Antivirals:

- Herpes
 - influenza
 - HIV

25 8. Antifungals and yeasts

- oral and vaginal candidiasis (e.g., glucomannan complex, α -D-MAN(1-6)_n branched α -(1-2) with L-RHAM, D-Gal,

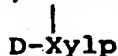
-23-



- actinomycetes

9. Food additives:

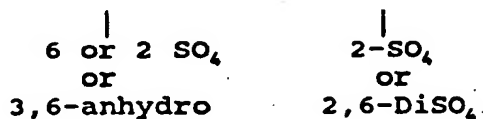
(e.g., Gum tragacanth, α -D-GalAp(1-4)_n


$$\pm \alpha\text{-L-Fucp or } \pm \beta\text{-D-Galp}$$

- emulsifiers

- thickeners

(e.g., carragenan (family), [D-Galp α -(1-3) D-Galp β -(1-4)]_n)



10. Veterinary:

- antibacterial
- antiviral
- antifungal
- anti-inflammatory

The present invention thus also provides pharmaceutical and other compositions, such as foodstuff compositions, containing saccharide compositions prepared in accordance with the present invention. In both the pharmaceutical compositions and the foodstuff compositions provided by the invention, the saccharide composition of the invention may be present in an amount of from $10^{-3} \mu\text{g ml}^{-1}$ to 100 mg ml^{-1} .

The concentration of the saccharide composition of

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the present invention in any given particular pharmaceutical composition or foodstuff composition will vary in terms of the activity of the saccharide being used. For pharmaceutical composition the concentration of saccharide present in the composition will depend on the in vitro activity measured for any given compound. For foodstuff compositions, the concentration of the saccharide composition of the present invention may be determined in accordance with the known activity of the compound being added.

For example, mothers milk contain the saccharide composition set forth above where it is indicated as being useful both in infant formula and as an antibacterial for fighting urinary tract infections. As such, the present invention provides an improvement in commercial infant formulas by permitting the addition to these commercial infant formulas the saccharide composition illustrated above. The particular saccharide composition illustrated above may be present in the commercial infant formula in an amount of 0.1 μg per ml to 1000 μg per ml. It is present in mother's milk at ca. 10 μg per ml.

The pharmaceutical compositions should be pyrogen free. Pharmaceutical compositions in accordance to the present invention may be prepared as is known in the art so as to be suitable for oral, intravenous, intramuscular, rectal, transdermal or nasal (e.g., nasal spray) administration. It may also be prepared for topical administration in the form of creams, ointments, suspensions, etc.

A few saccharides have been noted as being important both as commodity chemicals in the food, textile, and

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petroleum industries, and as specialty chemicals, primarily in the medical field, to date, the absence of an efficient process for preparing saccharide compositions has made it impossible to obtain commercial
5 compositions containing, as an active ingredient, a saccharide composition.

The present invention makes such saccharide compositions readily available in large quantity for the first time. With the method of the present invention,
10 saccharide compositions heretofore available only in miniscule quantities, and saccharide compositions heretofore unavailable, are readily made in gram and kilogram quantities. The purity of the saccharide compositions provided in accordance to the present
15 invention exceeds 95 wt.%. In some applications requiring high level of purity, the method of the present invention can be used to obtain saccharide compositions containing purity levels approaching 98 wt.% to essentially 100 wt.%.

20 The present invention thus now provides for the first time pharmaceutical compositions and other compositions containing saccharide compositions present in an effective amount. The present invention provides compositions containing the saccharide
25 compositions obtained in accordance with the present invention present in the amount of at least 100 mg, preferably at least 500 mg, and up to 95 wt.% of the composition.

In another embodiment, the present invention
30 provides an apparatus suitable for use in accordance with the present invention for the glycosyltransferase catalyzed synthesis of a saccharide composition.

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Illustrative configurations for such apparatus are :
provided in Figures 1, 2 and 3.

In a very basic embodiment the apparatus of the
present invention contains one reaction chamber in which
5 all of the glycosyltransferases, all the preselected
saccharide units and the initial acceptor moiety are
combined. Due to the specificity of the
glycosyltransferases, this mixture, given sufficient
time, will produce the saccharide composition of the
10 present invention.

Figures 1, 2 and 3 illustrate more efficiently
designed apparatuses which may be used in accordance with
the present invention. The apparatus illustrated in the
figures, comprise, as its basic elements, a reactor
15 equipped with an inlet and an outlet. The reactor is
suitable for carrying out the sequential covalent bonding
of a plurality of preselected saccharide units onto an
acceptor moiety, catalyzed by a plurality of
corresponding glycosyltransferases specific to each
20 covalent bonding. It contains at least three, preferably
four, and even more preferably a number greater than
four, such as five, six, seven, or more, different,
glycoltransferases which are preferably immobilized.

The inlet means is suitable for introducing the
25 acceptor moiety and the plurality of preselected
saccharide units into the reactor such that the
saccharide composition is synthesized. Preferably, the
inlet means is suitable for also introducing into the
reactor the glycosyltransferases which are themselves
30 preferably immobilized. The outlet means permits
discharging the saccharide composition from the reactor.

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Figure 1 illustrates a column-type reactor charged with a solid support matrix. The various glycosyltransferases (enzymes 1, 2, 3) used in the process may be either randomly distributed throughout the solid support matrix or they may be arranged in zones as illustrated in Figure 1. The initial acceptor moiety (shown as A in the figures) and the preselected saccharide units (shown as B, C and D in the figures) are charged into the reactor via the inlet means and passed through the solid support matrix whereupon the saccharide composition is produced due to the action of the specific glycosyltransferases and recovered via the outlet means as molecule A-B-C-D.

In the embodiment illustrated in Figure 2, the initial acceptor moiety and the preselected saccharide unit to be attached to the initial acceptor moiety are charged at the top of the solid support matrix, with the glycosyltransferases specific to the addition of each preselected saccharide units being arranged in corresponding zones along the direction of flow of the reaction mixture. The various preselected saccharide units are then individually added at correspondingly appropriate locations along the flow of the reaction mixture as shown in the figure.

In another preferred embodiment, illustrated in Figure 3, the reactor comprises a plurality of (n) reaction zones serially connected so as to be in sequential fluid communication with each other where (n) roughly corresponds to not more than the number of saccharide units being attached. Each reaction zone contains at least one glycosyltransferase specific to catalyze the bonding of a particular preselected saccharide unit onto the intermediate product formed in

-28-

the preceding reaction zone.

In accordance with this embodiment the initial acceptor moiety (A) and the first preselected saccharide unit (B) to be attached to the acceptor moiety are passed
5 through the first reaction zone which comprises a glycosyltransferase specific to catalyze the bonding of the first preselected saccharide unit onto the initial acceptor moiety thus producing a first intermediate product. This first intermediate product is then
10 transferred to the second reaction zone ($n=1$) where it is combined with the second preselected saccharide unit (X_n) and the glycosyltransferase ($E_{1,n}$) specific to catalyze the bonding of the second preselected saccharide unit with the first intermediate product formed. This process
15 is repeated in a corresponding number of reaction zones until the target saccharide composition illustrated as A-B-...-(X)- $_n$...Z, wherein each X moiety is independently selected, is obtained.

In another preferred embodiment, also illustrated in
20 Figure 3, means for purifying each intermediate product formed from the reaction mixture emanating from any given reaction zone is situated in fluid communication and between each of the reaction zones. The means for purifying removes contaminants in the reaction mixtures
25 which inhibit the efficiency of the bonding of the next preselected saccharide unit onto the intermediate product formed.

Additional objects, advantages, and novel features of this invention will become apparent to those skilled
30 in the art upon examination of the following examples thereof, which are not intended to be limiting.

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EXAMPLE 1Preparation of the Trisaccharide N-Acetylneuraminy 2-3 Galactosyl 8 1-4 N-Acetylglucosamine:

To each of five test tubes was added 10 μ l of pH 7.4
5 potassium phosphate buffer, 10 μ l of 50 mM MnCl_2 , 17,000
CPM of cytidine monophosphate-[^{14}C]-N-acetyneuraminic
acid, 25 μ l of galactosyltransferase, and 25 μ l of
N-acetyneuraminytransferase. The glycosyltransferases
were purified from bovine colostrum by Sephadex G-100 gel
10 chromatography.

To test tube 1 was also added 10 μ l of 40 mM uridine
diphosphate galactose and 10 μ l of 40 mM
N-acetylglucosamine. Test tube 1 was incubated in ice for
one hour.

15 To test tube 2 was also added 10 μ l of 40 mM uridine
diphosphate galactose. Test tube 2 was incubated at 37°C
for one hour.

To test tube 3 was also added 10 μ l of 40 mM
N-acetylglucosamine. Test tube 3 was incubated at 37°C
20 for one hour.

To test tubes 4 and 5 were also added 10 μ l of 40 mM
uridine diphosphate galactose and 10 μ l of 40 mM N--
acetylglucosamine. Test tubes 4 and 5 were incubated at
37°C for one hour.

25 After incubation, the contents of the test tubes
were each subjected to high voltage electrophoresis on
paper saturated with sodium tetraborate. Isotopically
labeled trisaccharide product was identified by its
mobility, as demonstrated by the product formed in test
30 tube 3.

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<u>Test Tube</u>	<u>Trisaccharide (cpm)</u>
1	0
5 2	0
3	3375
4	670
5	954

As can be seen, the presence of suitable acceptor moieties, donor moieties, and glycosyltransferases in test tubes 4 and 5 yielded the expected trisaccharide product from monosaccharide starting materials.

Typically, the sialic acid N-acetylneuraminate presents special problems for synthetic organic chemists seeking to incorporate it into saccharide compositions, due to the acid lability of its glycosidic bond. Synthesizing a trisaccharide from cytidine monophosphate N-acetylneuraminic acid enzymatically eliminates the synthetic problems associated with removing protecting groups under strong acidic condition.

It is believed that an acceptor moiety (N-acetylglucosamine) initially contacts a donor moiety (uridine diphosphate galactose) and a glycosyltransferase (galactosyltransferase) to produce a saccharide composition (galactosyl B 1-4 N-acetylglucosamine), which then acts as an acceptor moiety upon contacting a second donor moiety (cytidine monophosphate N-acetylneuraminic acid) and a second glycosyltransferase (N-acetylneuraminytransferase).

The synthesis of the trisaccharide product in test tubes 4 and 5 from monosaccharide starting materials is confirmed by comparison with the product of test tube 3, in which the trisaccharide is formed by contacting a

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disaccharide acceptor moiety (N-acetyllactosamine) with cytidine monophosphate Nacetylneuraminic acid and N-acetylneuraminyltransferase.

The absence of trisaccharide in test tube 2 illustrates that a suitable acceptor moiety is necessary for trisaccharide formation. The absence of trisaccharide in test tube 1 indicates that the synthesis of the trisaccharide is, indeed, dependent upon the action of any enzyme (the glycosyltransferase) that is inactive at low temperatures.

It is expected that the oligosaccharides N-acetylgalactosaminyl α 1-3 (fucosyl α 1-2) galactosyl β 1-4 Nacetylglucosaminyl β 1-3 galactose (a target for diarrheacausing bacteria) and N-acetylgalactosaminyl β 1-4 galactosyl β 1-4 glucose (a target for pneumonia-causing bacteria) can likewise be prepared by the processes of the present invention.

EXAMPLE 2

Tetrasaccharide biosynthesis protocol

Enzymes:

N-acetylglucosaminyltransferase:

Human colostrum is centrifuged for one hour at 70,000 x G. A 25% saturated ammonium sulfate cut yields a supernatant that is dialyzed to remove the ammonium sulfate. The retentate is applied to a Sephadex G-200 column (2.5 x 83 cm). The protein profile is determined spectrophotometrically at 280 nm, and a radioactive assay is performed to locate the fractions with transferase activity. The fractions containing the single enzyme peak are pooled and concentrated 10-fold by Amicon filtration. The pooled enzyme preparation is again assayed, and the protein concentration is determined

-32-

using a BioRad assay. The specific activity of the preparation is 5.3 pMoles per μ g protein-min.

Galactosyltransferase:

Human colostrum is centrifuged at 8700 x G for 15 minutes. The supernatant is poured through cheesecloth and 10 ml is applied to a Sephadex G-100 column (2.5 x 90 cm). The protein profile is determined spectrophotometrically at 280 nm, and a radioactive assay is performed to locate the fractions with enzyme activity. The fractions with the highest activity are pooled and concentrated 10-fold by Amicon filtration. The pooled enzyme preparation is again assayed, and the protein concentration is determined as above. The specific activity of the preparation is 15.4 pMoles per μ g protein-min.

Enzyme immobilization:

N-acetylglucosaminyltransferase:

300 mgs of Eupergit beads (1.2 ml) are washed three times with deionized water, and then three times with aseptic Hepes-buffered water. One ml of the enzyme preparation is combined aseptically with the beads along with UDP, lactose, $MnCl_2$, (final concentrations: 10, 25, and 10 mM, respectively) and a drop of chloroform in a Hepes-buffered solution. The beads are gently agitated at 4°C for 2½ days. Aliquots are taken and assayed periodically. To stop the derivatization, the beads are washed three times with an aseptic buffer, and stored in buffer, in the cold, with UDP, lactose, $MnCl_2$, and chloroform.

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Galactosyltransferase:

3.75 grams of beads are washed three times with deionized water, and then three times with aseptic Hepes-buffered water. The beads are added to 3-mls of the enzyme preparation (in both cases, optimum derivatization occurs at about 1 mg protein per 200 mgs beads) along with UDP, GlcNAc, $MnCl_2$, (final concentrations are all 10 mM) and a drop of chloroform in a Hepes-buffered solution. Derivatization and storage are as described above, except that the GlcNAc is used with the galactosyltransferase in place of lactose, which is the acceptor for the N-acetylglucosaminyltransferase.

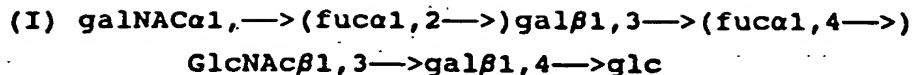
Tetrasaccharide production:

Derivatized N-acetylglucosaminyltransferase (0.5 ml beads) is incubated under constant stirring with lactose (25 mM), UDPGlcNAc (80 μM), and $MnCl_2$ (10 mM) for 21 hours. This incubation is carried out in duplicate-the supernatant of one incubation is used to measure the amount of trisaccharide produced (14 μg), and the supernatant from the other incubation is added to 0.5 ml beads derivatized with the galactosyltransferase. The galactosyltransferase incubation contains, therefore, 14 μg of trisaccharide, 25 μM UDPgal, and 10 mM $MnCl_2$. After 24 hours at room temperature, the second enzyme preparation produced about 1.6 μg of tetrasaccharide. After 31 hours, 2.2 μg of tetrasaccharide were produced.

EXAMPLE 3

The following schemes will be used for synthesizing three, relatively complex oligosaccharides: the A- and B-type milk oligosaccharides (I and II), and gum tragacanth (III), a plant oligosaccharide used by the ton as a food additive.

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First, the hexanolamine glycoside ($\text{glc-O-(CH}_2\text{)}_6\text{-NH}_2$) of glucose that will be affixed to CNBr-activated supports, e.g., Sepharose, via the amino group of the hexanolamine will be synthesized. Then the glucose-recognizing galactosyltransferase will be purified from human milk or colostrum using this affinity ligand. The enzyme, once partly purified, will be used to galactosylate glucose, making lactose.

Alternatively, the hexanolamine glycoside of lactose, which is an inexpensive and readily available disaccharide, will be synthesized. The lactose so produced will be attached to Sepharose and used as an affinity ligand will then be used to purify in part the N-acetylglucosaminyltransferase from human colostrum, or from human plasma.

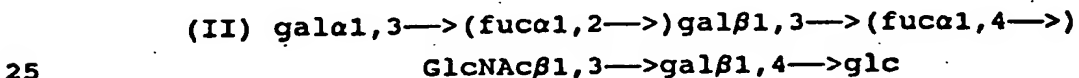
Next, this second transferase will be used to add N-acetylglucosamine to lactose, making the trisaccharide, which will again be attached to Sepharose. This bound trisaccharide will be used to obtain the $\beta 1, 3$ galactosyltransferase (from porcine submaxillary gland), which will, in turn, yield the substrate for purifying the next enzyme -- the $\alpha 1, 4$ fucosyltransferase (from porcine liver). The $\alpha 1, 2$ fucosyltransferase (from porcine submaxillary gland), and, finally, the $\alpha 1, 3$ N-acetylgalactosaminyltransferase (from porcine submaxillary glands) that terminates the synthesis of the A-type milk oligosaccharide will be affinity purified in this step-wise fashion. Each transferase so obtained will be immobilized to a solid matrix by any of several means, and the matrices will be poured in column

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configurations.

The enzyme-containing columns will be used sequentially, in the same order that the smaller amounts of derivatized substrates were synthesized, to synthesize large amounts each soluble oligosaccharide. A more tedious alternative method would be to use each enzyme in turn to synthesize a small amount of its product, which would then be immobilized onto Sepharose via a hexanolamine linker. This alternative method would require that hexanolamine linkers be attached to six compounds, and that derivitizations be accomplished with six hexanolamine-containing glycosides. The method requires only a single hexanolamine step, and a single derivitization of glucose-hexanolamine to Sepharose, which is a relatively uncomplicated process.

The order of attachment of the sugars is critical. The proximal fucose (that attached $\alpha 1,4$ to glcNAc) must be attached to the completed core tetrasaccharide before the addition of the second fucose (that attached $\alpha 1,2$ to the galactose. Finally, the terminal galNAc ($\alpha 1,3$) is added to complete the seven-sugar oligosaccharide. This order is required by the specificities of the glycosyltransferases.



Having synthesized I, II will be synthesized in precisely the same fashion, except that the hexasaccharide will be used, first, to purify an $\alpha 1,3$ galactosyltransferase that will be derivitized with protective groups for a galactosyl-, and not an N-acetylgalactosyltransferase. This enzyme will then be

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used to synthesize the B-type oligosaccharide.

(III) [...(fucal,3→xylβ1,3→)galA
α1,4→(gal,β1,4→xylβ1,3→)galA...]

5 To isolate the enzyme that synthesizes the α1,4
galacturonic acid backbone of gum tragacanth, which
currently is available only from the bark of a tree
species indigenous to the Middle East, hexagalacturonans
will be prepared from pectin, a common constituent of
citrus rinds, and used as an affinity ligand.

10 The same affinity ligand can be used, next to
isolate, from tree tissues, the xylosyltransferase that
synthesizes the proximal β1,3 xylosides. The xylosylated
galacturonans, once derivatized, will be used to isolate
both the fucosyl- and galactosyltransferases that,
15 respectively, fucosylate and galactosylate the
xylosylated galacturonan. In the case of this
oligosaccharide, the degree of xylosylation,
fucosylation, and galactosylation will be controlled
empirically by the number of passes of the compounds
20 through the appropriate enzyme-containing columns. The
number of repeat units produced will depend on the number
of galacturonic acid residues used initially; this number
will vary in length from four to twenty monosaccharide
units.

25

* * * * *

Obviously, numerous modifications and variations of
the present invention are possible in light of the above
teachings. It is therefore to be understood that within
the scope of the appended claims, the invention may be
30 practiced otherwise than as specifically described
herein.

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Claims

1. A method for the glycosyltransferase-catalyzed preparation of a saccharide composition by serially bonding preselected saccharide units to an acceptor moiety, comprising the steps of:

(i) preparing a glycosyltransferase capable of transferring a preselected saccharide unit to an acceptor moiety by contacting said acceptor moiety with a mixture suspected to contain a plurality of glycosyltransferases under conditions effective to bind said acceptor moiety and said glycosyltransferase, and thereby isolating said glycosyltransferase, wherein said acceptor moiety is one member selected from the group consisting of proteins, glycoproteins, lipids, glycolipids, and carbohydrates;

(ii) providing conditions and co-reagents sufficient to effect bonding of said preselected saccharide unit to said acceptor moiety catalyzed by said glycosyltransferase thereby obtaining a product; and

(iii) performing steps (i) and (ii) a plurality of times such that the product obtained in step (ii) of a given iteration is used as the acceptor moiety in step (i) of the following iteration until said saccharide composition is obtained.

2. The method of Claim 1 wherein said carbohydrate is one member selected from the group consisting of monosaccharides, disaccharides, oligosaccharides, and polysaccharides.

3. The method of Claim 1 wherein the glycosyltransferase used in step (ii) is immobilized to a solid support.

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4. The method of Claim 3 wherein said glycosyltransferase attached to a solid support is obtained by protecting the active site of said glycosyltransferase during the immobilization process.

5 5. The method of Claim 1 wherein said co-reagents comprise manganese cations.

6. The method of Claim 1 wherein said saccharide used in step (ii) is a saccharide nucleotide.

10 7. The method of Claim 5 wherein said nucleotide is one member selected from the group consisting of uridine, guanosine and cytidine phosphates.

15 8. The method of Claim 1 wherein said acceptor moiety used in said first iteration is one member selected from the group consisting of proteins, glycoproteins, lipids, and glycolipids.

9. The method of Claim 1 wherein said acceptor moiety used in said first iteration is one member selected from the group consisting of monosaccharides, disaccharides, oligosaccharides, and polysaccharides.

20 10. The method of Claim 1 wherein said acceptor moiety used in said first iteration is N-acetylglucosamine.

25 11. The method of Claim 1 wherein said acceptor moiety used in said first iteration is N-acetylglucosamine, said glycosyltransferase used in said first iteration is galactosyltransferase, and said glycosyltransferase used in said second iteration is N-

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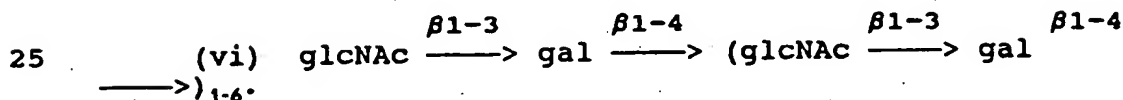
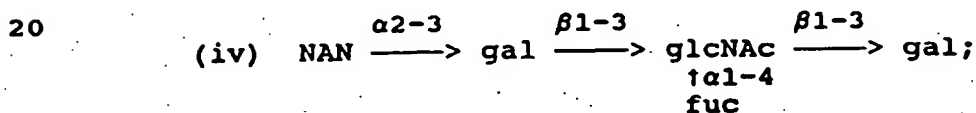
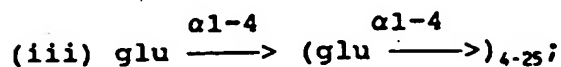
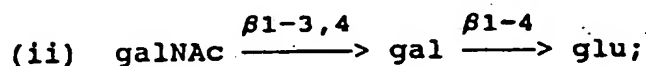
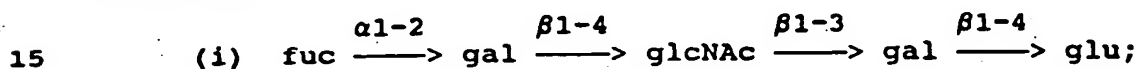
acetylneuraminyltransferase.

12. The method of Claim 1 wherein said acceptor moiety used in said second iteration is galactosyl B 1-4 N-acetylglucosamine.

5 13. The method of Claim 1 wherein at least one of the donor moieties used is cytidine monophosphate N-acetylneuraminic acid.

10 14. The method of Claim 1 wherein said mixture suspected to contain a plurality of glycosyltransferases is a cell homogenate.

15 15. The method of Claim 1 wherein said saccharide composition is a compound of one of the following formulae:



16. A pharmaceutical composition comprising, in association with pharmaceutically acceptable excipient or carrier, a saccharide composition other than heparin

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prepared by a method for glycosyltransferase-catalyzed serial bonding of preselected saccharide units to an acceptor moiety comprising the steps of:

(i) preparing a glycosyltransferase capable of
5 transferring a preselected saccharide unit to an acceptor moiety by contacting said acceptor moiety with a mixture suspected to contain said glycosyltransferase under conditions effective to bind said acceptor moiety and said glycosyltransferase, thereby isolating said
10 glycosyltransferase, wherein said acceptor moiety is one member selected from the group consisting of proteins, glycoproteins, lipids, glycolipids, and carbohydrates;

(ii) providing conditions and co-reagents
sufficient to effect bonding of said preselected
15 saccharide unit to said acceptor moiety catalyzed by said glycosyltransferase thereby obtaining a product; and

(iii) performing steps (i) and (ii) a plurality of times such that the product obtained in step (ii) of a given iteration is used as the acceptor moiety of step
20 (i) of the following iteration.

17. The pharmaceutical composition of Claim 16 comprising at least 100 mg of said saccharide composition.

18. The pharmaceutical composition of Claim 16
25 comprising at least 1 gram of said saccharide composition.

19. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a protein.

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20. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a glycoprotein.

21. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a lipid.

22. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a glycolipid.

23. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a carbohydrate.

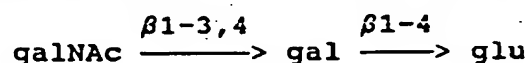
24. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a monosaccharide.

25. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a disaccharide.

26. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is an oligosaccharide.

27. The pharmaceutical composition of Claim 16 wherein said acceptor moiety used in said first iteration is a polysaccharide.

28. A pharmaceutical composition suitable for use in the therapy or treatment of pneumonia comprising an effective amount of a compound of the formula:



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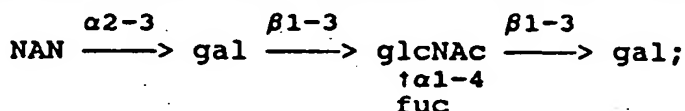
in association with a pharmaceutically acceptable carrier or excipient.

29. A pharmaceutical composition suitable for use in the therapy or treatment of periodontal disease comprising an effective amount of a compound of the formula:



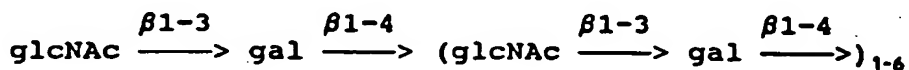
in association with a pharmaceutically acceptable carrier or excipient.

30. A pharmaceutical composition suitable for use in the therapy or treatment of solid tumors comprising an effective amount of a compound of the formula:



in association with a pharmaceutically acceptable carrier or excipient.

31. A pharmaceutically composition suitable for use as a contraceptive comprising an effective amount of a compound comprising the formula:



- in association with a pharmaceutically acceptable carrier or excipient.

32. A foodstuff comprising a saccharide composition

other than gum tragacanth or carrageenan prepared by a method for glycosyltransferase-catalyzed serial bonding of preselected saccharide units to an acceptor moiety comprising the steps of:

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(i) preparing a glycosyltransferase capable of transferring a preselected saccharide unit to an acceptor moiety by contacting said acceptor moiety with a mixture suspected to contain a plurality of glycosyltransferases under conditions effective to bind said acceptor moiety and said glycosyltransferase, and thereby isolating said glycosyltransferase, wherein said acceptor moiety is one member selected from the group consisting of proteins, glycoproteins, lipids, glycolipids, and carbohydrates;

(ii) providing conditions and co-reagents sufficient to effect bonding of said preselected saccharide unit to said acceptor moiety catalyzed by said glycosyltransferase thereby obtaining a product; and

(iii) performing steps (i) and (ii) a plurality of times such that the product obtained in step (ii) of a given iteration is used as the acceptor moiety in step (i) of the following iteration.

33. In an infant foodstuff, the improvement comprising a compound of the formula:

(i)
$$\text{fuc} \xrightarrow{\alpha 1-2} \text{gal} \xrightarrow{\beta 1-4} \text{glcNAc} \xrightarrow{\beta 1-3} \text{gal} \xrightarrow{\beta 1-4} \text{glu}$$

present in an amount of from $0.1 \mu\text{g ml}^{-1}$ to $10 \mu\text{g ml}^{-1}$.

34. An apparatus for the glycosyltransferase-catalyzed synthesis of a saccharide composition, said apparatus comprising:

a reactor;

at least four different glycosyltransferases in said reactor;

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inlet means for introducing an acceptor moiety and a plurality of preselected saccharide units into said reactor such that said saccharide composition is synthesized; and

- 5 outlet means for discharging said saccharide composition from said reactor;

 wherein said acceptor moiety is one member selected from the group consisting of proteins, glycoproteins, lipids, glycolipids, and carbohydrates.

- 10 35. The apparatus of Claim 34 wherein said reactor comprises a plurality of reaction zones serially connected so as to be in sequential fluid communication with each other, wherein each reaction zone comprises a glycosyltransferase specific to catalyze bonding of a
15 preselected saccharide unit onto the intermediate product formed in the preceding reaction zone.

36. The apparatus of Claim 34 wherein means for purifying the intermediate products formed in each of said reaction zones from the rest of the reaction mixture
20 produced therein is situated in fluid communication and between each of said reaction zones.

 37. The apparatus of Claim 34 wherein said glycosyltransferases are immobilized onto a solid support.

- 25 38. The apparatus of Claim 37 wherein the active sites of said glycosyltransferases immobilized onto said solid support are protected during the immobilization process.

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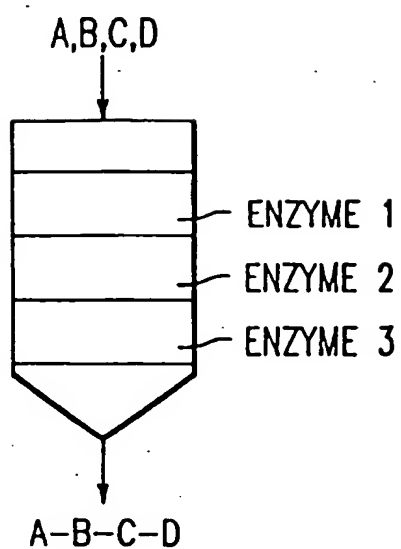


FIG. 1

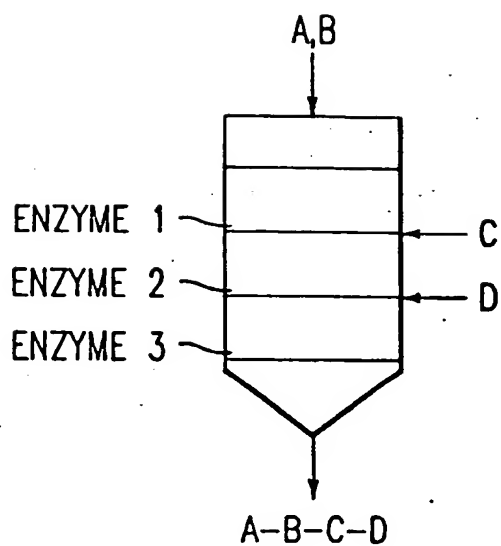


FIG. 2

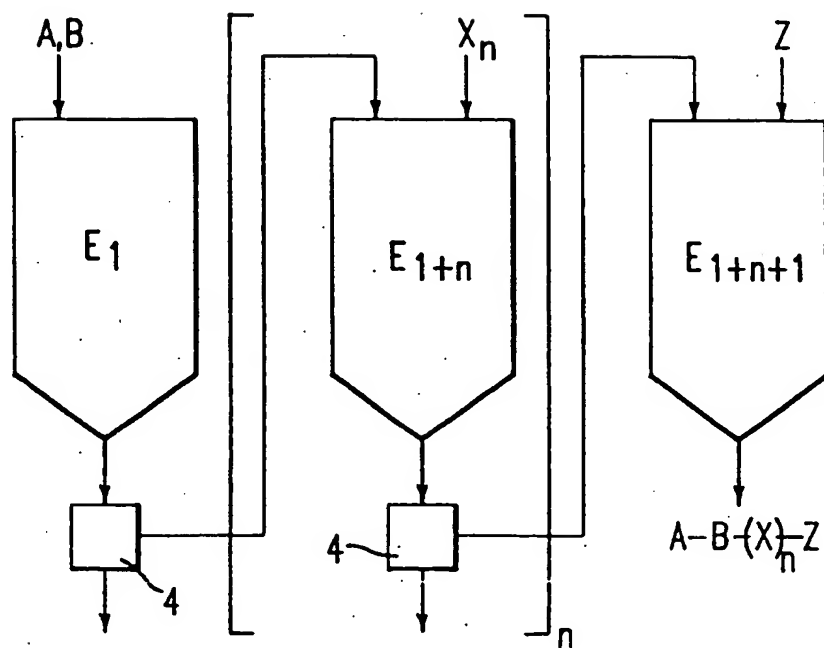


FIG. 3

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